

1 Roger S. Borovoy (# 031209)
2 David J. Miclean (# 115098)
3 Kurtis D. MacFerrin (# 178006)
4 Tamara Fraizer (#215492)
5 FISH & RICHARDSON P.C.
6 500 Arguello Street, Suite 500
7 Redwood City, CA 94063
8 Telephone: (650) 839-5070
9 Facsimile: (650) 839-5071
10 Attorneys for Defendant and Counter-Claimant
11 MOLECULAR DEVICES CORPORATION

UNITED STATES DISTRICT COURT
NORTHERN DISTRICT OF CALIFORNIA
SAN FRANCISCO DIVISION

12 CALIPER TECHNOLOGIES CORP.,
13 Plaintiff,
14 v.
15 MOLECULAR DEVICES CORPORATION,
16 Defendant.

Case No. C 02-01837 JSW (JL)

MOLECULAR DEVICES' OPPOSITION TO MOTION FOR PRELIMINARY INJUNCTION

[REDACTED]

Date: May 16, 2003
Time: 9:00 A.M.
Ctrm: 2

Judge: Honorable Jeffrey S. White

AND RELATED COUNTERCLAIMS.

**MOLECULAR DEVICES' OPPOSITION TO MOTION FOR
PRELIMINARY INJUNCTION [REDACTED]**
Case No. C 02-01837 JSW (JL)

1 TABLE OF CONTENTS

2	I.	INTRODUCTION	1
3	II.	SUMMARY OF ARGUMENT	2
4	III.	STATEMENT OF FACTS	3
5	A.	Kinase Enzyme Research.....	3
6	B.	Fluorescence Polarization Is An Old Technique.....	4
7	C.	The Prior Art Fluorescence Polarization Kinase Assay.....	5
8	D.	Caliper Failed to Disclose Material Prior Art (The Seethala Reference) to the U.S. Patent and Trademark Office.....	7
9			
10			
11			
12	H.	Caliper's Delay.....	13
13	I.	Caliper Is Not Being Irreparably Harmed By Sales Of IMAP™ Assays	14
14	J.	MDC Would Be Substantially Harmed By An Injunction	16
15	IV.	ARGUMENT	17
16	A.	A Preliminary Injunction Is A Harsh Remedy.....	17
17	B.	Caliper Is Not Likely To Succeed At Trial	18
18	1.	The IMAP™ assays do not infringe the '774 and '141 patents	18
19	a.	IMAP™ assays do not use "non-specific, charge dependent" binding	19
20	b.	IMAP™ assays do not include a "comparing" or "monitoring" step	20
21	c.	Caliper has not shown that MDC uses IMAP™ assays to infringe the patents, or induces or contributes to infringement by others	22
22	2.	The Caliper patents are invalid and unenforceable	22
23	C.	Caliper Will Not Be Irreparably Harmed In The Absence Of A Preliminary Injunction	24
24			
25			
26			
27			
28			

1	1.	Caliper Is Not Entitled To A Presumption Of Irreparable Harm.....	24
2	2.	Caliper Unreasonably Delayed in Seeking Injunctive Relief	25
3	3.	Caliper's Licensing of the Patents-in-Suit Negates Any Claim Of Irreparable Harm	26
4	4.	Caliper Fails To Show How Its Reputation And Market Share Are Affected By MDC.....	27
5	D.	The Hardship That MDC Will Suffer If The Preliminary Injunction Is Granted Outweighs Any Potential Harm To Caliper	29
6	E.	The Public Interest Dictates Against Granting Injunctive Relief	29
7	V.	CONCLUSION.....	30

10
11
12
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15
16
17
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3	<i>Aro Mfg. Co. v. Convertible Top Replacement Co.</i> , 365 U.S. 336 (1961).....	18
4		
5	<i>Associated Gen. Contractors of Cal. v. Coalition for Econ. Equity</i> , 950 F.2d 1401 (9th Cir. 1991).....	24
6		
7	<i>Atari Corp. v. Sega of Am., Inc.</i> , 869 F. Supp. 783 (N.D. Cal. 1994)	24, 27
8		
9	<i>Campbell Soup Co. v. ConAgra, Inc.</i> , 977 F.2d 86 (3d Cir. 1992).....	24
10		
11	<i>Caribbean Marine Serv. Co. v. Baldrige</i> , 844 F.2d 668 (9th Cir. 1988).....	24
12		
13	<i>Datascope Corp. v. Kontron Inc.</i> , 786 F.2d 398 (Fed. Cir. 1986).....	29
14		
15	<i>Elk Corp. of Dallas v. GAF Bldg. Materials Corp.</i> , 168 F.3d 28 (Fed. Cir. 1999).....	23
16		
17	<i>EMI Group N. Am., Inc. v. Intel Corp.</i> , 157 F.3d 887 (Fed. Cir. 1998).....	18
18		
19	<i>FMC Corp. v. United States</i> , 3 F.3d 424 (Fed. Cir. 1993).....	24
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22		
23	<i>High Tech Med. Instrumentation, Inc. v. New Image Indus., Inc.</i> , 49 F.3d 1551 (Fed. Cir. 1995).....	17, 24, 26, 27
24		
25	<i>Ill. Tool Works, Inc. v. Grip-Pak, Inc.</i> , 906 F.2d 679 (Fed. Cir. 1990).....	17, 30
26		
27	<i>Intel Corp. v. ULSI Sys. Technology</i> , 995 F.2d 1566 (1993).....	16, 17
28		
29	<i>Johnson & Johnston Assocs. Inc. v. R.E. Serv. Co., Inc.</i> , 285 F.3d 1046 (Fed. Cir. 2002).....	18
30		
31	<i>LaBounty Mfg., Inc. v. U.S. Int'l Trade Comm'n</i> , 958 F.2d 1066 (Fed. Cir. 1992).....	23
32		
33	<i>Novartis Corp. v. Ben Venue Labs., Inc.</i> , 271 F.3d 1043 (Fed. Cir. 2001).....	18
34		
35	<i>Nutrition 21 v. United States</i> , 930 F.2d 867 (Fed. Cir. 1991).....	17, 24
36		

1	<i>Playboy Enters., Inc. v. Netscape Communications Corp.</i> , 55 F. Supp. 2d 1070 (C.D. Cal. 1999).....	25
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10	<i>Stokely-Van Camp, Inc. v. Coca-Cola Co.</i> , No. 86-C-6159, 1987 U.S. Dist. LEXIS 781 (N.D. Ill. Jan. 30, 1987)	25
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13	<i>Telemac Cellular Corp. v. Topp Telecom, Inc.</i> , 247 F.3d 1316 (Fed. Cir. 2001).....	18
14	<i>Vehicular Techs. Corp. v. Titan Wheel Inc.</i> , 141 F.3d 1084 (Fed. Cir. 1998).....	17, 24
16	<u>Federal Statutes</u>	
17	35 U.S.C. § 102(a)	23
18	35 U.S.C. § 112	19, 22
19	<u>Other Authorities</u>	
20	11 Charles A. Wright & Arthur R. Miller, Federal Practice and Procedure § 2948 (1969)	17
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1 **I. INTRODUCTION**

2 Caliper Technologies Corp. ("Caliper") is not entitled to a preliminary injunction against
3 Molecular Devices Corporation ("MDC") after delaying over a year to bring its motion, and
4 because *MDC does not use the fluorescence polarization assay method of the '774 and '141*
5 *patents in MDC's IMAP™ assay kits.* In fact, not even Caliper uses the patented method.

6 Caliper was well aware of MDC's IMAP™ technology in September 2001, and even
7 obtained IMAP™ assay kits when they were released by MDC a few months later. Yet, Caliper
8 waited months before filing suit, and many more months before bringing this motion.

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12 By bringing this action, Caliper appears to be trying to prevent the use of the better assay
13 (IMAP™) that runs on customers' existing equipment so that Caliper can sell its more expensive,
14 highly specialized, but often slow and inconvenient equipment. There can be no other reason, for
15 Caliper does not even sell assay kits using its own patented technique. Caliper's motion for
16 preliminary injunction should be denied.

17 **II. SUMMARY OF ARGUMENT**

18 The preliminary injunction motion should be denied because:

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- 3 • Caliper's '774 and '141 patent claims are invalid as they are anticipated by the Seethala
4 publication, and are unenforceable for failure to disclose material prior art (Seethala) to the U.S.
5 Patent and Trademark Office.
- 6 • Caliper cannot establish irreparable harm; where it delayed bringing the instant injunction
7 motion over a year after it knew of MDC's IMAP™ technology, and obtained MDC's IMAP™
8 assay kits; and where Caliper's failure to sell customers on its new high-throughput screening,
9 "systems" has less to do with IMAP™, and more to do with the fact that Caliper's system
10 doesn't work well, doesn't measure fluorescence polarization, and is a departure from industry
11 standards.
- 12 • Further, the harm to MDC's business reputation, customer relationships and vendor contracts
13 should an injunction issue, far outweighs any hardship Caliper would experience while waiting
14 for a remedy after a verdict in this case.
- 15 • Public policy also favors continuing drug research without interruption.

16 **III. STATEMENT OF FACTS**

17 **A. Kinase Enzyme Research**

18 The MDC IMAP™ assay kits at issue in this case are designed to detect and measure the
19 activity of an enzyme, particularly a kinase enzyme, using an old technique called fluorescence
20 polarization. Kinases are enzymes that occur naturally in the body and are essential for the normal
21 function of cells. Abnormal kinase activity has been linked to various cancers, auto-immune
22 diseases, inflammatory diseases (such as asthma), diabetes, neurological disorders and
23 cardiovascular (heart) diseases. (See Declaration of J. Richard Sportsman ("Sportsman Decl.")
24 ¶ 12.) Thus, there is considerable interest in finding drugs that "inhibit" kinase activity ("kinase
25 inhibitors"), which could be used to control these diseases.

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1 Although there are many different kinases in the body, *each kinase does one simple thing –*
2 *it transfers a phosphate from one molecule to another.*⁶ Detecting whether that transfer of
3 phosphate takes place is what IMAP™ and other kinase assays are all about. If the kinase inhibitor
4 drug being tested has no effect on the kinase (the drug doesn't work), the phosphate will be
5 transferred by the kinase (the kinase did work). If the drug being tested works to inhibit the kinase,
6 the phosphate will not be transferred by the kinase. (Sportsman Decl. ¶¶ 13, 14 and 44.) This
7 process of the kinase transferring a phosphate from one molecule to another is called
8 “phosphorylation”. If the assay results in a “phosphorylated” product, it means the kinase did its
9 job and was not inhibited – that is, the drug didn't work. *Id.* In drug research, the kinase usually
10 works because finding an inhibiting drug is like finding the needle in the haystack.

11 **B. Fluorescence Polarization Is An Old Technique.**

12 Fluorescence polarization is a detection technique that has been known since the 1920's
13 when scientists first developed the techniques to measure and analyze it. (Sportsman Decl. ¶ 11.)
14 Fluorescence is similar to what makes glow-in-the-dark toys and posters visible after you shine light
15 on them. Things that fluoresce emit light shortly after being “hit” with light. Scientists can attach a
16 molecule that emits fluorescent light – known as a fluorescent “tag” or “label” – to a molecule that
17 does not emit fluorescent light, and which would otherwise be invisible. Scientists can then detect
18 the otherwise invisible molecule by the fluorescent “glow” of the tag that was attached to it.
19 (Sportsman Decl. ¶ 7.)⁷

20 Fluorescence polarization measures the “polarization” of the light emitted by the
21 fluorescently labeled tag on the molecule (polarization is essentially light emitted along one
22 direction or plane). (Sportsman Decl. ¶ 8.) Small fluorescently tagged molecules tumble and spin
23 quickly emitting light in many random directions. The net result is “low fluorescence polarization”.
24 Conversely, large, slow moving fluorescently tagged molecules tend to emit light in one plane or
25 direction – this is “high fluorescence polarization”. (Sportsman Decl. ¶¶ 7-11.) Accordingly, the

26 ⁶ Phosphate is labeled as PO₄ because it has one phosphorous atom and four oxygen atoms. (Sportsman
27 Decl. ¶ 13.)

28 ⁷ A fluorescently tagged molecule can be analogized to a small glow-in-the-dark sticker affixed to a golf ball
in a dark room. You could not see the golf ball, but you could see the sticker attached to it.

1 fluorescence polarization of molecules can be used to determine whether there are large or small
2 fluorescent molecules in the sample or mixture. (Sportsman Decl. ¶ 9.) Small fluorescent
3 molecules will have low fluorescence polarization because they spin and tumble quickly, while
4 large, slow molecules will have high fluorescence polarization.

5 Fluorescence polarization has also been used to determine whether a small fluorescently
6 tagged molecule is bound to a bigger molecule. When the small molecule is bound to a large
7 molecule, it "acts and looks" like a large molecule. That is, it tumbles slowly and has high
8 fluorescence polarization. (Sportsman Decl. ¶ 10.) If the small molecule does not bind to the large
9 one, it acts and looks like the small molecule, that is, tumbling fast and having a low fluorescence
10 polarization. Thus, measuring the fluorescence polarization is one way to determine whether the
11 small molecule is of a kind that could bind to a particular large one. Low fluorescence polarization
12 indicates that the small molecule did not bind with a big one, while high fluorescence polarization
13 indicates that the small molecule did bind to the large one.⁸

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22 **C. The Prior Art Fluorescence Polarization Kinase Assay**

23 Like the earlier fluorescence polarization methods, the Caliper method in the '774 and '141
24 patents is used to see if a kinase was able to transform a "starting material" into what is called the
25 "reaction product", by transferring a phosphate group to the reaction product. (Miclean Decl. Ex. P;

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27 ⁸ Binding a fluorescently tagged small molecule to a large molecule can be analogized to attaching the golf
28 ball with its glow-in-the-dark sticker, to a large, heavy medicine ball, which would dramatically slow
down the movement of the golf ball (if the golf ball were spinning).

1 Nikiforov Tr. 265:20-266:6; 140:5-18; 140:23-142:18; 271:15-24; Barron Decl. ¶ 6). The starting
2 material has two important features: (1) it has a fluorescent label or tag attached to it, which will
3 allow scientists to track what happens to it, and (2) it has the ability to have a phosphate group added
4 to it by the kinase.⁹ In both the Caliper and the earlier fluorescence polarization methods, this
5 fluorescently tagged starting material is placed into a mixture containing water, among other
6 things.¹⁰ Then, ATP¹¹ and the kinase (which has been incubated with the potentially inhibitory
7 drug) are added to that mixture.¹² If the kinase works (and is not inhibited), it will transfer a
8 phosphate group from ATP to the fluorescently-labeled starting material, creating the fluorescently
9 tagged reaction product – a “phosphorylated” version of the starting material.¹³ (Miclean Decl. Ex.
10 P; Nikiforov Tr. 26:12-15; 43:7-25; 93:20-94:3; 265; 271:21-272:10; 272:15-21; Barron Decl.
11 Ex. C.)

12 Yet, regardless whether the kinase worked to transfer a phosphate to the starting material –
13 thus creating the reaction product – both the fluorescently tagged starting material and reaction
14 product would be small molecules that spin fast and tumble – emitting their light in many directions
15 (low fluorescence polarization.) This would make them indistinguishable. To detect and
16 distinguish the reaction product (to determine if the kinase worked) scientists had to make it act like
17 a big and slow molecule so it could emit light in one direction (high fluorescence polarization.)
18 (Sportsman Decl. ¶ 8-10.)

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21 ⁹ In the words of claim 1 of the ‘774 patent, this is a “first reagent having a fluorescent label”; In the words
22 of claim 1 of the ‘141 patent, this is a “phosphorylatable compound that includes a fluorescent label”.
23 “Phosphorylatable” simply means that a phosphate group can be chemically added to the molecule by the
24 action of the kinase; that transferring of the phosphate by the kinase is called “phosphorylation”, and the
product of that reaction is a “phosphorylated” product. (Miclean Decl. Ex. MM, Col. 29, l. 1-25; Ex.
NN, Col. 37, l. 15-30 (Sportsman Decl.
¶ 13.).

25 ¹⁰ The Caliper patent claims call this mixture a “first mixture”.

26 ¹¹ The Caliper patent claims call ATP a “second reagent” or a “phosphate donor group”. ATP has three
phosphates attached to it. The kinase will take one of the ATP phosphates and give it to the starting
material.

27 ¹² Claim 1 of the ‘774 patent calls this new mixture a “second mixture”.

28 ¹³ Claim 1 of the ‘774 patent calls the product a “fluorescently labeled product having a substantially
different charge than the first reagent” and the ‘141 patent calls it a “phosphorylated product.”

When the reaction product binds to the “binding reagent”, it will be slowed down allowing its fluorescent tag to emit light in one direction (high fluorescence polarization.) Scientists could then detect the high fluorescence polarization and know the kinase did its job, (and the drug did not work.) Conversely, if the kinase was inhibited and there was no reaction product created (no phosphate transferred by the kinase) there would be no binding with the binding reagent and only low fluorescence polarization.¹⁵

A publication by Seethala et al. (“Seethala”) described exactly this fluorescence polarization method for assaying kinase activity, and did so before Nikiforov came up with his invention.

Barron Decl. ¶ 5, Ex. C). Seethala's binding reagent was an antibody.¹⁶

D. Caliper Failed to Disclose Material Prior Art (The Seethala Reference) to the U.S. Patent and Trademark Office

Seethala describes a kinase assay that uses fluorescence polarization in the same way that the Caliper method uses it. (Barron Decl. ¶¶ 5-7, 12, and 34.) It is therefore

¹⁴ Claim 1 of the '774 patent calls the binding reagent a "polyionic polymer" and claim 1 of the '141 patent calls it a "polycationic component".

¹⁵ Using the golf ball analogy, the starting material would be the golf ball with the glow-in-the-dark sticker. The reaction product would be the golf ball and sticker with an added phosphate group transferred by the activity of the kinase (if the kinase was not inhibited by the drug.) To slow down the reaction product and to make it distinguishable from any starting material, scientists would bind it to a large heavy molecule "the binding reagent"; like binding a large heavy medicine ball to the golf ball with sticker and added phosphate. This would slow the golf ball (reaction product) down and make it act more like the medicine ball, which would reduce the spin and tumble, and result in a more polarized light emission from glow-in-the-dark sticker.

¹⁶ Seethala's antibody, like all antibodies, was a protein, and all proteins are polymers of amino acids. A "polymer" is made up of repeating units. Proteins are polymers made up of a chain of units called amino acids (which include tyrosine). Depending on the surrounding conditions, some of the amino acids in the antibody will have a positive electrical charge, some a negative charge, and others no charge. (Barron Decl. 5, Ex. C)

1 material prior art that was known to the inventor and to Caliper, and it should have been disclosed
2 by them to the U.S. Patent and Trademark Office. It was not.

3 E.

4 The only change Dr. Nikiforov made to the Seethala fluorescence polarization method for
5 kinase assays was to use a binding reagent that was not an antibody

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²⁰ Attached as Exhibit MM to the Miclean declaration.
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H. Caliper's Delay

On September 11, 2001, the '774 patent issued to Caliper. At this time, MDC had already released its first IMAP™ kit and was working to finalize production of its first IMAP™ kinase assay kits, which Caliper now accuses of infringing its '774 patent. (Sportsman Decl. ¶¶ 35-37, 41-42.) Caliper admits to knowing about MDC's work "in the fall of 2001." (Merion Supp. Decl. to Caliper's Mot. Prelim Inj. ¶ 16; Despite knowing

about MDC's IMAP™ products (and even buying and testing three kits)²⁴ Caliper sent no "cease and desist" communications to MDC before filing suit, and delayed suing MDC for seven months after the '774 patent issued. Further, Caliper did not move for a preliminary injunction on the '774 patent at the time it filed the action in April of 2002. (Miclean Decl. Ex. S (Caliper's Complaint).)

The second Caliper patent, the '141 patent, which Caliper acknowledges "describes essentially the same method [as the '774 patent] in a slightly different way," (Caliper Br. at 15:24-25), issued October 29, 2002. While Caliper now tries to explain that the issuance of the '141 patent was delayed because of clerical errors in the patent office, the errors were actually Caliper's, and Caliper did nothing to follow up on the delay or mitigate it until after the application for the

²⁴ Caliper purchased a "PDE" IMAP™ kit in August 2001 and another in November 2001; Caliper purchased an SGK IMAP™ kit in February 2002. (Miclean Decl. Ex. KK, Ex. P; Nikiforov Tr. 72:22-76:22.)

1 '141 patent was abandoned. (See ft. 34, *infra*.) Once the '141 patent was issued in October 2002,
2 Caliper amended its complaint to allege infringement, but, again, did not at that time move for a
3 preliminary injunction on either patent. (Miclean Decl. Ex. T (Caliper's First Am. Complaint).) It
4 was not until 16 months after Caliper's first patent issued, and the IMAP™ technology was
5 disclosed by MDC to the world in September 2001, that Caliper brought this preliminary injunction
6 motion.

I. Caliper Is Not Being Irreparably Harmed By Sales Of IMAP™ Assays

8 Caliper claims that its market position in high-throughput screening kinase assays is being
9 irreparably harmed by MDC's sale of IMAP™ assays. (Caliper Br. at 3-4, 21-24.) This claim is
10 without basis. Initially, Caliper does not offer a fluorescence polarization kinase assay kit (such as
11 IMAP™) for sale. Instead, it sells a system that

12 uses a mobility shift assay technique—not the fluorescence polarization technique of the patents-in-
13 suit

17 Decl. ¶¶ 42-44.) Thus, any claims or intimation that MDC has taken sales away from Caliper by
18 selling the IMAP™ kinase assay kits are, at best, grossly overstated, and, at worst, false.

19 Further, Caliper has not used the technology described in the patents-in-suit in a commercial
20 product, nor does it appear to have any plans to do so in the future. (Caliper Br. at 4, fn. 2;

Instead of standardized microplates, Caliper's methods use what they call a "LabChip"—a device having very small grooves, through which solutions are moved. (Oldfield Decl. ¶ 15; Miclean Decl. Ex. W.) Experiments on LabChips are read by special instruments that are marketed only by Caliper, and

27 Thus, even if

1 fluorescence polarization kinase assays could be done on LabChips, the results could not be seen
2 using Caliper's LabChip equipment.

3 In addition, kinase assays are not the new, emerging field that Caliper makes them out to be.
4 Well over *half a billion* kinase assays are done every year. (Oldfield Decl. ¶ 9, Ex. N; Sportsman
5 Decl. ¶¶ 11-12.) As of November of 2000, nearly a year before the first of the patents-in-suit
6 issued, there were at least ten companies offering twenty-eight different non-radioactive kinase
7 assay kits. (Oldfield Decl. ¶ 10, Ex. O (The Scientist 14[22]:20, Nov. 13, 2000, and accompanying
8 chart; Sportsman Decl. ¶ 23.) These long-available kits even include approaches that do not use
9 antibodies, and do use fluorescent tags. *Id.* Currently, there are about twenty companies offering
10 about thirty kinase assays, about a third of which are non-radioactive/non-antibody.

11 All of
12 these many available kits are run on various existing high-throughput instruments. *Id.*

13 Caliper's claim of irreparable harm is based solely on the declaration of Michael Merion,
14 plaintiff's Vice President of Sales. This declaration expounds on Dr. Merion's view of the kinase
15 assay market and IMAP™'s supposed impact on Caliper's sales. But Dr. Merion's declaration is
16 without foundation. In sharp contrast to both the statements and the implications in his declaration,
17 in deposition

Dr. Merion admitted:

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5 Accordingly, Caliper's claims of irreparable harm should be viewed with much skepticism.

6 **J. MDC Would Be Substantially Harmed By An Injunction**

7 Caliper and MDC are both small, publicly traded companies. Caliper's motion for a
8 preliminary injunction threatens to completely shut down MDC's IMAP™ business

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19 The issuance of a preliminary injunction would place MDC at a great disadvantage both in the
20 marketplace and in this litigation.

21 **IV. ARGUMENT**

22 **A. A Preliminary Injunction Is A Harsh Remedy**

23 As Caliper correctly observes, the law of the Federal Circuit governs the substantive
24 standards underlying the grant or denial of preliminary injunctions in patent cases. *Hybritech, Inc.*
25 *v. Abbott Labs*, 849 F.2d 1446, 1451 n.12 (Fed. Cir. 1988). The Federal Circuit on many occasions
26 has cautioned that a preliminary injunction is a discretionary tool providing a drastic and
27 extraordinary remedy that is not to be routinely granted. *See, e.g., Intel Corp. v. ULSI Sys. Tech.*,

1 *Inc.*, 995 F.2d 1566, 1568 (Fed. Cir. 1993); *Ill. Tool Works, Inc. v. Grip-Pak, Inc.*, 906 F.2d 679,
2 683 (Fed. Cir. 1990) (citing 11 Charles A. Wright & Arthur R. Miller, *Federal Practice and*
3 *Procedure* § 2948 (1969)). A party seeking a preliminary injunction must meet a well-established
4 four-factor inquiry: (1) that it has a reasonable likelihood of success on the merits; (2) that it will
5 suffer irreparable harm if preliminary relief is not granted; (3) that the balance of its hardships
6 against the hardships of the party against whom the injunction is sought weighs in its favor; and (4)
7 that the public interest is not adversely impacted. *Vehicular Techs. Corp. v. Titan Wheel Inc.*, 141
8 F.3d 1084, 1087-88 (Fed. Cir. 1998) and *Intel Corp. v. ULSI Sys. Technology*, 995 F.2d 1566
9 (1993).

10 The burden to show entitlement to a preliminary injunction is not light and rests always on
11 the moving party. *Reebok Int'l Ltd. v. J. Baker, Inc.*, 32 F.3d 1552, 1555 (Fed. Cir. 1994)*8.
12 Central to the movant's burden are the likelihood of success and the irreparable harm factors.
13 *Sofamor Danek Group, Inc. v. DePuy-Motech, Inc.*, 74 F.3d 1216, 1219 (Fed. Cir. 1996). A movant
14 in a patent infringement suit, must first show a reasonable likelihood of success on the merits that
15 requires proof of both patent validity and infringement. *Id.* Specifically, “because of the
16 extraordinary nature of the relief, the *patentee* carries the burden of showing likelihood of success
17 on the merits with respect to the patent's validity, enforceability, and infringement.” *Nutrition 21 v.*
18 *United States*, 930 F.2d 867, 869 (Fed. Cir. 1991) (emphasis original). “In other words, if [the
19 accused infringer] raises a ‘substantial question’ concerning validity, enforceability, or infringement
20 (i.e., asserts a defense that [the patentee] cannot show ‘lacks substantial merit’) the preliminary
21 injunction should not issue.” *Genentech, Inc. v. Novo Nordisk*, 108 F.3d 1361, 1364 (Fed. Cir.
22 1997). Where there is no clear showing of a likelihood of success in proving infringement, there is
23 no presumption of irreparable harm. *High Tech Med. Instrumentation, Inc. v. New Image Indus.,*
24 *Inc.*, 49 F.3d 1551, 1556 (Fed. Cir. 1995). In balancing the hardships, a weak showing of likelihood
25 of success tips the balance in favor of the party against whom the injunction is sought. *Ill. Tool*
26 *Works, supra*, 906 F.2d at 683.

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1 **B. Caliper Is Not Likely To Succeed At Trial**

2 Caliper is not likely to succeed at trial, because the IMAP™ assays do not infringe the
3 Caliper patent claims, and because those claims are invalid.

4 **1. The IMAP™ assays do not infringe the '774 and '141 patents**

5 In patent law, the scope of the protected invention is defined by the claims at the end of the
6 patent. *Aro Mfg. Co. v. Convertible Top Replacement Co.*, 365 U.S. 336, 339 (1961) (“[T]he claims
7 made in the patent are the sole measure of the grant . . .”); *Johnson & Johnston Assocs. Inc. v. R.E.*
8 *Serv. Co., Inc.*, 285 F.3d 1046, 1052 (Fed. Cir. 2002) (“Both the Supreme Court and this court have
9 adhered to the fundamental principle that claims define the scope of patent protection.”). Caliper
10 has the burden of proving that the claims are infringed. *Novartis Corp. v. Ben Venue Labs., Inc.*,
11 271 F.3d 1043, 1046 (Fed. Cir. 2001). Whether or not a claim is infringed is determined in two
12 steps: first, words and phrases in the claim are interpreted to determine the proper scope of the
13 claim; second, the claim as interpreted is compared with the accused product or method to see if the
14 product or method includes each and every element of the claims. *Telemac Cellular Corp. v. Topp*
15 *Telecom, Inc.*, 247 F.3d 1316, 1323 (Fed. Cir. 2001). Here, Claim 1 of the '774 and '141 patents,
16 the only claims at issue in this motion, do not cover a product, they cover methods.²⁵ Thus, the
17 IMAP™ assay kits sold by MDC cannot infringe the claims by themselves, contrary to Caliper's
18 suggestions otherwise. (Caliper Br. at 1:1-11, 11:15-16.) Rather, the claims are only infringed if
19 someone performs each and every step of the claimed methods using the IMAP™ assays. *EMI*
20 *Group N. Am., Inc. v. Intel Corp.*, 157 F.3d 887, 896 (Fed. Cir. 1998).

21 a.

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²⁵ Claim 1 of the '774 patent and claim 1 of the '141 patent are the only claims at issue in Caliper's motion.
28 (Caliper Br., p. 12:5-10.) Moreover, Caliper only discusses literal infringement, and does not apply the
 doctrine of equivalents. Thus, this brief addresses only literal infringement of those two claims.

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25 ²⁶ Miclean Decl. Ex. II [Caliper Prelim Claim Constructions], at p. 2:20-27 (construing "polyionic polymer"
26 (the binding reagent) in claim 1 of the '774 patent), p. 9:3-11 (construing "polycationic component,"(the
27 binding reagent in the '141 patent). For purposes of analyzing infringement, MDC has assumed that
28 Caliper is correct in contending that the claims should be interpreted this way. However, MDC reserves
 the right to argue that they should not, and are therefore invalid under 35 U.S.C. § 112, as discussed
 below at fn. 30.

27 ²⁷ Claims must be construed the same way for purposes of determining infringement as they are for
28 determining validity. *See Amazon.com, Inc. v. BarnesandNoble.com, Inc.*, 239 F.3d 1343, 1351 (Fed. Cir.
 2001).

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b. IMAP™ assays do not include a “comparing” or “monitoring” step

Claim 1 of the '774 patent requires the step of “comparing a fluorescent polarization in the second mixture relative to the first mixture” (Miclean Decl. Ex. MM, Col. 29, Claim 1.)

This “comparing” step is, therefore, completely lacking from the instructions that accompany the IMAP™ assay kits. (Barron Decl. ¶¶ 29-30, 33.)

1 Claim 1 of the '141 patent requires the step of "monitoring a level of phosphorylated
2 product produced by the activity of the kinase enzyme" (Miclean Decl. Ex. NN, Col. 37, l 25-
3 30.) The concept of "monitoring" was added to the '141 patent application (the '774 patent does
4 not use the term) to describe multiple measurements over time of the fluorescence polarization of a
5 reaction mixture.²⁸ As already shown, the IMAP™ assays do not measure a *difference* in
6 fluorescence polarization – nor do they make other multiple measurements of fluorescence
7 polarization.

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12 Although there are still more reasons why the IMAP™ assays do not infringe the claims of
13 the '774 and '141 patents, for the sake of brevity those reasons are not presented here

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16 c. **Caliper has not shown that MDC uses IMAP™ assays to infringe
17 the patents, or induces or contributes to infringement by others**

18 MDC cannot be held liable for infringement unless it performs the claimed methods itself
19 (direct infringement), or unless it induces or contributes to infringement by others (indirect

20 ²⁸ The term "monitoring" and related terms are first referenced in the '141 patent in Example 3, "*Time
21 Course Monitoring of Enzyme Reactions by Fluorescence Polarization*". (Miclean Decl. Ex. NN; '141
22 patent at Column 27:1-26 (emphasis added)). In that example, "assays were *monitored over time* in order to
determine the efficacy of the fluorescence polarization detection methods of the present invention on
23 *monitoring reaction time courses*" (emphasis added). Example 4 explains that "[t]he fluorescent
polarization detection methods of the present invention were also applied in *monitoring the time course* of a
24 phosphatase assay." *Id.* at Column 27:27-45. In that example, "[t]he relative fluorescence polarization level
was *monitored over time* for a control reaction (no enzyme) and a reaction mixture with different
concentrations of phosphatase enzyme." *Id.* (emphasis added). As Example 7 explains, "[t]hermal
25 denaturation experiments were also performed in the presence of polylysine, while *monitoring fluorescence
polarization changes* in the reaction mixtures." ('141 patent at Column 29:44-46 (emphasis added)). *In
sum, the '141 patent consistently uses the term "monitoring" only to describe multiple fluorescence
polarization measurements of the same sample.* See *Bell Atl. Network Servs., Inc. v. Covad
Communications Group, Inc.*, 262 F.3d 1258, 1268, 1270-71 (Fed. Cir. 2001) ("[S]pecification may define
claim terms 'by implication,' and 'when a patentee uses a claim term throughout the entire patent
specification, in a manner consistent with only a single meaning, he has defined that term 'by implication.'"
The court found that the ordinary meaning of the disputed non-technical term "mode" was "sufficiently
broad and amorphous that the scope of the claim language can be reconciled only with recourse to the written
description.").

1 infringement). Even if it was possible to infringe the Caliper patent claims using the IMAP™
2 assays (which it is not), Caliper has failed to present sufficient evidence that MDC itself uses the
3 IMAP™ assays in an infringing manner, or that MDC induces or contributes to such infringement
4 by others.²⁹

5 **2. The Caliper patents are invalid and unenforceable**

6 There also remains substantial questions concerning the validity and enforceability of the
7 ‘774 and ‘141 patent claims in view of the prior art.³⁰ The only modification that Dr. Nikiforov’s
8 alleged invention makes to the prior art Seethala assay, is to use a non-antibody peptide as a binding
9 reagent, instead of the antibody used in the Seethala assay. Indeed, Caliper does not dispute that
10 every element of Claim 1 of the ‘774 and ‘141 patents, other than the particular binding reagent that
11 they used, was disclosed in Seethala’s prior art publication.³¹ (See comparison diagrams attached as
12 Ex. I to Barron Decl. ¶ 34.)

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15 ²⁹ Caliper simply asserts, with no evidence, that MDC uses the IMAP™ assays to provide product
16 demonstrations and support to customers. (Caliper Br. at 12:11-12). But Caliper has not established how
17 MDC or anyone else has actually used the IMAP™ assays. Caliper has not met its burden of proving that
18 MDC should be held liable because it contributed to or induced any infringement. A plaintiff asserting
19 contributory infringement must prove four elements: (1) the defendant sold a component or material for use
20 in practicing the patented method; (2) the component or material constitutes a material part of the invention;
21 (3) the defendant knew that the item it sold was especially made or adapted for use in infringing the patented
22 method; and (4) the item sold is not a staple article or commodity of commerce suitable for substantial
23 noninfringing use. *See C.R. Bard, Inc. v. Advanced Cardiovascular Sys., Inc.*, 911 F.2d 670, 673 (Fed. Cir.
24 1990). In order to establish inducement of infringement, it must be shown that the alleged inducer
25 knowingly and actively induced infringement. *Warner-Lambert Co. v. Apotex Corp.*, 316 F.3d 1348, 1363
26 (Fed. Cir. 2003). It must be established that the defendant possessed specific intent to encourage another’s
27 infringement and not merely that the defendant had knowledge of the acts alleged to constitute inducement.
Warner-Lambert, 316 F.3d at 1363. The plaintiff has the burden of showing that the alleged infringer’s
28 actions induced infringing acts and that he knew or should have known his actions would induce actual
infringements. *Id.*; *see also Hewlett-Packard Co. v. Bausch & Lomb Inc.*, 909 F.2d 1464, 1468-69 (Fed. Cir.
1990) (discussing statutory development of the doctrine of active inducement of infringement). Caliper has
not attempted to prove each and every one of these elements, and in particular has made no effort to show
that the IMAP™ assays are not capable of substantial non-infringing uses or that MDC had the specific
intent to encourage infringement by others.

29 As discussed in Molecular Devices’ Preliminary Invalidity Contentions, the claims of the ‘774 and ‘141
30 patents fail to meet several requirements of 35 U.S.C. § 112, in particular the written description
requirement. (See Miclean Decl. Ex. X (MDC’s Prelim Invalid. Cont. and Supp.).)

31 They are therefore invalid.
See *Gentry Gallery, Inc. v. Berkline Corp.*, 134 F.3d 1473, 1479 (Fed. Cir. 1998) (finding claims invalid on
written description grounds because the claims omitted “an essential element” of the invention disclosed in
the original disclosure, and therefore impermissibly broadened the scope of the claims). For the sake of
brevity, MDC’s other invalidity arguments will not be discussed at greater length in this brief.

32 Barron Decl. ¶ 34, Ex. I. There are many other publications that also disclose some or all of the elements
of the claims, but for the sake of brevity those other references will not be discussed in this brief.

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11 every element of the claims. (Barron Decl. ¶ 34.) *See In re Paulsen*, 30 F.3d 1475, 1478-79 (Fed.
12 Cir. 1994) (“A rejection for anticipation under section 102 requires that each and every limitation of
13 the claimed invention be disclosed in a single prior art reference.”). Indeed, the comparison
14 diagrams of Exhibit I to the declaration of Barron, a slightly modified version of Exhibit E to
15 Caliper’s brief, shows how the prior art Seethala assay has each and every element of Caliper’s
16 claims thus anticipating (and invalidating) the Caliper patents.

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19 Such non-disclosure of
20 material prior art constitutes inequitable conduct sufficient to render the patents unenforceable.
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23 ³² An antibody is a polymer of amino acids, with negative and positive charges as a result of those amino
24 acids. (Miclean Decl. Ex. P; Nikiforov Tr. 159:15-160:1; 181:11-21.)

25 ³³ Caliper’s failure to disclose the Seethala publication is one basis for MDC’S position that the claims are
26 unenforceable due to Caliper’s inequitable conduct in failing to disclose material prior art to the United
27 States Patent and Trademark Office (“the Patent Office”). (Miclean Decl. Ex. AA.) (*See also Elk Corp.
of Dallas v. GAF Bldg. Materials Corp.*, 168 F.3d 28, 30 (Fed. Cir. 1999) (noting that a breach of patent
applicants’ duty to prosecute patent applications in the Patent Office “with candor, good faith, and
honesty . . . may constitute inequitable conduct”); *LaBounty Mfg., Inc. v. U.S. Int’l Trade Comm’n*, 958
F.2d 1066, 1074, 1076 (Fed. Cir. 1992) (finding undisclosed prior art reference material for inequitable
conduct analysis and noting that “[p]atent applicants are required to disclose to the examiner all
information ‘material to the examination of the application’”). For the sake of brevity, Caliper’s
inequitable conduct will not be discussed at greater length in this motion.

1 **C. Caliper Will Not Be Irreparably Harmed In The Absence Of A Preliminary**
2 **Injunction**

3 Parties seeking pretrial injunctive relief must demonstrate they will be exposed to some
4 irreparable harm if such relief is denied. *Vehicular Techs. Corp. v. Titan Wheel Int'l, Inc.*, 141 F.3d
5 1084, 1088 (Fed. Cir. 1998); *Associated Gen. Contractors of Cal. v. Coalition for Econ. Equity*,
6 950 F.2d 1401, 1410 (9th Cir. 1991). The threatened harm must be immediate and significant.
7 *FMC Corp. v. United States*, 3 F.3d 424, 430 (Fed. Cir. 1993) (finding that plaintiff's failure to
8 establish the extent of the irreparable harm precluded a grant of preliminary injunction); *Caribbean*
9 *Marine Serv. Co. v. Baldridge*, 844 F.2d 668, 674 (9th Cir. 1988). To establish "irreparable injury",
10 Caliper must demonstrate potential harm that cannot be redressed by a legal or equitable remedy
11 following trial. The preliminary injunction must be the only way of protecting the plaintiff from
12 such harm. *Campbell Soup Co. v. ConAgra, Inc.*, 977 F.2d 86, 91 (3d Cir. 1992). Caliper has not
13 shown irreparable harm in the instant motion—its over year delay in seeking a preliminary
14 injunction alone shows that the extraordinary relief of an injunction before a full trial on the merits
15 is unwarranted.

16 **1. Caliper Is Not Entitled To A Presumption Of Irreparable Harm**

17 While a patentee may be entitled to a presumption of irreparable harm when it makes a
18 strong showing of likelihood of success on the merits and a showing of continuing infringement, see
19 *Reebok Int'l Ltd. v. J. Baker, Inc.*, 32 F.3d 1552, 1556 (Fed. Cir. 1994), there is no such
20 presumption where, as here, there has been no strong showing of likelihood of success. See *Atari*
21 *Corp. v. Sega of Am., Inc.*, 869 F. Supp. 783, 790 (N.D. Cal. 1994). MDC has demonstrated that
22 Caliper is unlikely to succeed on the merits as to both infringement and validity. Caliper has not,
23 therefore, established entitlement to a presumption of irreparable harm.

24 **2. Caliper Unreasonably Delayed in Seeking Injunctive Relief**

25 Caliper's claim of irreparable harm is belied by its over 16-month delay in seeking to enjoin
26 MDC'S development and sale of IMAP™ assays. "[D]elay in seeking a remedy is an important
27 factor bearing on the need for a preliminary injunction. *High Tech Med. Instrumentation, Inc.*, 49
28 F.3d at 1557; see also *Nutrition 21 v. United States*, 930 F.2d 867, 872 (Fed. Cir. 1991) (Patentee's

1 substantial delay before seeking a preliminary injunction “suggests that the status quo does not
2 irreparably damage [the patentee].”); *T.J. Smith & Nephew Ltd.*, 821 F.2d 646, 648 (1987) (noting
3 that presumption of irreparable harm would have been rebutted by a fifteen-month delay in seeking
4 an injunction in combination with the grant of licenses).

5 Caliper has moved for a preliminary injunction a full sixteen months after its first patent
6 issued and the MDC IMAP™ technology was disclosed to the world in September 2001. Caliper’s
7 motion simply comes too late. *Playboy Enters., Inc. v. Netscape Communications Corp.*, 55 F.
8 Supp. 2d 1070, 1080, 1090 (C.D. Cal. 1999) (5-month delay after retaining the expert in filing
9 preliminary injunction motion supported denial of motion); *Stokely-Van Camp, Inc. v. Coca-Cola*
10 Co., No. 86-C-6159, 1987 U.S. Dist. LEXIS 781, at *7 (N.D. Ill. Jan. 30, 1987) (“[T]he fact that
11 [plaintiff] waited three months indicates a lack of a need for the extraordinary remedy of a
12 preliminary injunction.”); see also, *Programmed Tax Sys., Inc. v. Raytheon Co.*, 419 F. Supp. 1251,
13 1255 (S.D.N.Y. 1976).

14 Caliper has no excuse for its failure to file a preliminary injunction motion for 16 months
15 after issuance of the ’774 patent. To the extent Caliper claims the delay in filing its preliminary
16 injunction motion was related to the delay in the issuance of the ’141 patent, such argument is
17 subterfuge. Initially, there is no reason Caliper could not have brought its motion based solely on
18 the ’774 patent, which, according to Caliper, is “essentially the same” as the ’141 patent. (Caliper
19 Br. at 15:24-26.) Further, delay in the issuance of the ’141 patent was related to Caliper’s
20 inattention and neglect.³⁴ Caliper’s claim that it had hoped to “prosecute this lawsuit in a
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24 ³⁴ Caliper argues that clerical errors in the patent office delayed the issuance of the ’141 patent. (Caliper Br.
25 at 20:9-13.) But it was Caliper, not the patent office, that was responsible for any delay. Caliper
26 received a notice of the allowance of its claims on January 2, 2002, and paid its issuance fee on January
27 10, 2002. Then, on March 1, Caliper filed a continuation application in order to try and change the
28 claims. But Caliper used the wrong procedure for the continuation and failed to withdraw the previous
claims. The new claims were allowed on March 8, 2002, but, because of Caliper’s mistake, the fee that
Caliper had already paid was never applied to the new application. More than five months later the PTO
issued a notice of abandonment. Only then did Caliper act, explaining its mistakes and petitioning for
the withdrawal of this holding. (Miclean Decl. Ex. BB (August 28, 2002 Pet. Withdraw Holding
Aband.).) The petition was approved, and the patent eventually issued on October 29, 2002 – almost 8
full months after it was allowed. Caliper’s procedural mistakes caused the delay, and that Caliper did not
follow up on this delay can only be its own fault.

1 reasonable period of time without the need for a preliminary injunction proceeding" should also be
2 viewed with skepticism where, as here, the case is being litigated within the normal time frame.³⁵

3 Moreover, Caliper's veiled accusations that MDC has been delinquent in its discovery
4 obligations, thus excusing Caliper's delay, are not only false, they are illogical. MDC's document
5 production is substantially complete and it was completed in timely fashion (despite the fact that it
6 was five times the size of Caliper's production.) (Miclean Decl. ¶ 14.) Fundamentally, though,
7 MDC's discovery responses have nothing to do with Caliper's determination of the supposed harm
8 that it is suffering.

9 **3. Caliper's Licensing of the Patents-in-Suit Negates Any Claim Of**
10 **Irreparable Harm**

11 A patentee's willingness to license its patent to competitors shows that money is adequate
12 compensation. *See High Tech Med. Instrumentation*, 49 F.3d at 1557; *see also Rosemount, Inc. v.*
13 *ITC*, 910 F.2d 819, 821-22 (Fed. Cir. 1990); *T. J. Smith & Nephew*, 821 F.2d at 648. If money
14 damages are sufficient, there is no irreparable harm because "any injury suffered by [the patentee]
15 would be compensable in damages assessed as part of the final judgment in the case." *High Tech*
16 *Med. Instrumentation*, 49 F.3d at 1557.

17 Contrary to the implication of Caliper's opening brief and the supporting Merion Decl.
18 ¶ 14), Caliper *does* license its patents, including to Agilent

19 Indeed, it is Caliper's *stated corporate policy* to license its technology. For instance, in
20 its corporate fact sheet, Caliper explains that it has two business models—direct sales and OEM
21 business. (Miclean Decl. Ex. U, Corporate Summary, p. 1.) Caliper further explains that "[t]he
22 OEM business provides chips and enabling technologies to vendors who then integrate the
23 application solution and market it to their end customers. Caliper's partnership with Agilent
24 Technologies, Inc. is a prime example of this latter approach." Id. That licensing is a focus of
25 Caliper is supported by its public financial documents. In 2002, out of total revenues of less than

26 ³⁵The average time to trial in this district in 2002 was 23.5 months, even in non-patent cases. (Miclean Decl.
27 Ex. CC (Judicial Caseload Profile, available on <http://www.uscourts.gov/cgi-bin/cmsd2002.pl>.) Even
28 with the short, three-month delay caused by Caliper adding the '141 patent, this is consistent with the

1 \$26M, over \$9M are from license fees and contract revenue. (Miclean Decl. Ex. GG.) Caliper's
2 willingness—indeed, desire—to license its technology indicates that money is sufficient to
3 compensate it for the use of its technology. A preliminary injunction is therefore unwarranted.

4 **4. Caliper Fails To Show How Its Reputation And Market Share Are
5 Affected By MDC.**

6 Caliper claims that its market position in high-throughput screening kinase assays is being
7 harmed by MDC's alleged infringement. (Caliper Br. at 3-4, 21-24.) But, as discussed above,
8 Caliper does not appear to currently offer a kinase assay kit (like IMAP™) for sale. Nor has
9 Caliper used the technology described in the patents-in-suit in a commercial product,

10 Finally,
11 kinase assays are a well-developed field with many competitors and technologies available.
12 (Oldfield Decl. ¶¶ 9-10.)

13 Caliper offers no evidence that MDC'S IMAP™ products have had, or can have, any effect
14 on Caliper's market share for kinase assays. Caliper has not identified any MDC customers that
15 would purchase kinase assays from Caliper, as opposed to its many competitors, if the injunction
16 issues. (See *infra*, p. 15, l. 8-28.) Also, MDC'S sales account for only about one percent of the
17 total market for kinase assays. (Oldfield Decl. ¶ 9.)

18 Moreover, the fact that Caliper does not use the assay technology described in the patents-
19 in-suit is a factor that weighs against a finding of irreparable harm. *Atari, supra*, 869 F. Supp. at
20 790-92. *See also Reiffin v. Microsoft Corp.*, 158 F. Supp. 2d 1016, 1028 (N.D. Cal. 2001) ("Given
21 the fact that [the plaintiff] does not manufacture, sell or license any products based on his patents,
22 he cannot point to any specific commercial interest that needs equitable protection.") (citing *High*
23 *Tech Med. Instrumentation*, 49 F.3d at 1556 ("[T]he lack of commercial activity by the patentee is a
24 significant factor in the calculus.")).

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28 current trial date, which is 21 months after filing. Thus, Caliper knew, or at least should have known, the
approximate time frame to trial that it could expect in this case.

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10 The instruments and the assays made by MDC and other companies use standard
11 microplates.³⁶ Because they are standardized, the instruments and the assays are cross-compatible.
12 That is, you can use any assay on any microplate, and read it with almost any instrument. Industry
13 data indicates that standard microplates make up the vast majority of all high-throughput screening.
14 (Oldfield Decl. ¶ 13.) Caliper, however, has chosen to market a different, purportedly more
15 “sophisticated” kind of plate and reader. Caliper’s HTS systems have also been publicly criticized
16 for use with enzymatic assays (like kinase enzyme assays) (Oldfield Decl. ¶¶ 13-15), and are slower
17 and more expensive than other HTS systems. (Oldfield Decl. ¶ 15-16.) Caliper’s failure in the
18 marketplace has nothing to do with its patents or MDC, and everything to do with the risky choice
19 to market something new that customers don’t want. (Oldfield Decl. ¶¶ 13-16; Miclean Decl. Ex.
20 PP.)

21 **D. The Hardship That MDC Will Suffer If The Preliminary Injunction Is Granted
22 Outweighs Any Potential Harm To Caliper**

23 A preliminary injunction is a harsh remedy that here essentially gives Caliper its trial
24 remedy before discovery has even been completed. Caliper’s motion for a preliminary injunction
25 threatens to completely shut down MDC’S IMAP™ business.

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28 ³⁶ Microplates are somewhat like a grid of tiny test wells attached together. These microplates come in
multiple sizes that allow companies to do from 6-3456 experiments at the same time.

Injunctions are truly

10 drastic remedies. They can often compel capitulation of a defendant regardless of the merits of its
11 defense. While MDC vehemently disputes Caliper's infringement accusations and the validity of
12 the patents-in-suit, an injunction may well make it prohibitive to litigate these issues. The hardship
13 to MDC of an injunction outweighs any theoretical harm to Caliper caused by denial of its motion.

E. The Public Interest Dictates Against Granting Injunctive Relief

15 The scientists who use IMAP™ assays are doing important research on cancer, autoimmune
16 diseases, inflammatory diseases (such as asthma), diabetes, and neurological and cardiovascular
17 (heart) disorders. (Oldfield Decl. ¶ 8.) If a preliminary injunction issues, these researchers would
18 be forced to switch assay approaches mid-stream.

19 Further, the legitimate public interest in competition and continuing critical research without
20 interruption favors the denial of the motion for preliminary injunction. While there always exists a
21 public interest in protecting the rights secured by a valid patent, the focus of the public interest
22 analysis should be whether there exists some public interest that would be injured by the grant of
23 the preliminary injunction. *Hybritech Inc. v. Abbott Labs.*, 849 F.2d 1446, 1458 (Fed. Cir. 1988);
24 *see also Datascope Corp. v. Kontron Inc.*, 786 F.2d 398, 401 (Fed. Cir. 1986) (affirming district
25 court's finding that public interest supported a denial of preliminary injunction because the accused
26 infringer had "made some showing that the public will be harmed by an injunction"). The interest

1 in protection of patent rights can be counterbalanced by the interest in competition. *See Ill. Tool
2 Works, Inc. v. Grip-Pak, Inc.*, 906 F.2d 679, 684 (Fed. Cir. 1990).

3 **V. CONCLUSION**

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13 There is, therefore, no likelihood of success on the merits, and Caliper's
14 motion for preliminary injunction should be denied.

15 Dated: April 1, 2003

FISH & RICHARDSON P.C.

16

17

By:


David J. McLean

18

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Attorneys for Defendant and Counter-Claimant
MOLECULAR DEVICES CORPORATION

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1 Roger S. Borovoy (# 031209)
2 David J. Miclean (# 115098)
2 Kurtis D. MacFerrin (# 178006)
3 Tamara Fraizer (#215492)
3 FISH & RICHARDSON P.C.
4 500 Arguello Street, Suite 500
4 Redwood City, CA 94063
Telephone: (650) 839-5070
5 Facsimile: (650) 839-5071

6 Attorneys for Defendant and Counter-Claimant
MOLECULAR DEVICES CORPORATION

7

8 UNITED STATES DISTRICT COURT

9 NORTHERN DISTRICT OF CALIFORNIA

10 SAN FRANCISCO DIVISION

11 CALIPER TECHNOLOGIES CORP.,

Case No. C 02-01837 JSW (JL)

12 Plaintiff,

13 v.

14 MOLECULAR DEVICES CORPORATION,

DECLARATION OF ANDREW R.
BARRON IN SUPPORT OF MOLECULAR
DEVICES' OPPOSITION TO MOTION
FOR PRELIMINARY INJUNCTION

15 Defendant.

[REDACTED VERSION]

16 Date: May 16, 2003

Time: 9:00 A.M.

Ctrm: 2

17 Judge: Honorable Jeffrey S. White

18
19 AND RELATED COUNTERCLAIMS.

20
21 I, Andrew R. Barron, declare and state:

22 1. I have been retained by Fish & Richardson P.C. to offer my expert opinion concerning
23 technical issues in this case. If called as a witness, I could competently testify to the following
24 facts, all of which are based on my own personal knowledge and expertise.

25 My Expertise and Preparation

26 2. I am currently the Charles W. Duncan Jr. – Welch Chair of Chemistry and Professor of
27 Materials Science at Rice University, Houston, TX.

28

1 3. I am an expert in chemistry and materials science, with particular experience in the area
2 of the Group 13 elements including gallium. I have been performing independent research in this
3 area for approximately seventeen years. My particular expertise with respect to the case includes
4 extensive experience with the reaction and structural characterization of gallium compounds,
5 especially the reactions with phosphate and related compounds. Evidence of this expertise is the
6 receipt of two awards from the Royal Society of Chemistry, the first being the Meldola Medal and
7 Prize in 1990, and in 1995, I was awarded the Corday Morgan Medal and Prize, both for my
8 research of gallium chemistry. I have published over 250 peer-reviewed papers, 5 book chapters,
9 and am a named inventor in approximately 12 patents. I have published a number of papers directly
10 relating to gallium phosphate chemistry. I have consulted for a number of US and foreign
11 companies in the area of gallium chemistry. Further references, a publication list, and related
12 documents are given in my *curriculum vitae*, see attached Exhibit A.

13 4. In considering the aspects of this case and formulating my opinions expressed herein, I
14 have used my knowledge and expertise of chemistry. In addition, I have used my knowledge
15 gained over years of studying general scientific texts and specialist journal articles, attending
16 scientific conferences, and engaging in scholarly discussions with colleagues. In particular in this
17 case, I have reviewed a number of papers, discovery documents, deposition transcripts, patents, and
18 I have had discussions with MDC personnel. I have relied on specific documents that are
19 referenced throughout this text. A complete list of the materials I have reviewed in preparing this
20 declaration is attached as Exhibit B. My work is continuing, however, and I may supplement this
21 declaration if additional information is produced by Caliper and provided to me. Further, my
22 testimony may contain additional information that I receive or gain between now and that time. At
23 the hearing, if called to testify, I expect to use demonstrative exhibits, including any Figures from
24 my report, but I have not yet decided on them specifically.

25 **Previously Known Kinase Assay Methods Using Fluorescence Polarization**

26 5. Caliper wasn't the first to use fluorescence polarization in a kinase assay. Kinase assays
27 have been accomplished previously using fluorescence polarization, as described in a publication by
28

1 Seethala and Menzel, "A Homogenous, Fluorescence Polarization Assay for Src-Family Tyrosine
2 Kinases," *Analytical Biochemistry*, 253, 210-218 (1997), attached as Exhibit C ("Seethala").

3 6. Seethala described the use of an antibody as a binding reagent to affect the fluorescence
4 polarization of phosphotyrosine. The phosphotyrosine was the product of a kinase reaction. The
5 phosphotyrosine is equivalent to the phosphoserine disclosed in Examples 1 and 2 in Caliper's U.S.
6 Patent No., 6,287,774 ("the '774 patent"), col. 23, line 60 to col. 24, line 67, and Examples 1-3 in
7 Caliper's U.S. Patent No. 6,472,141 ("the '141 patent"), col. 25, line 26 to col. 27, line 26.

8 7. As described in a later publication by Wilkinson, "Tyr'd and True: Immunochemical
9 reagents and kits for studying tyrosine phosphorylation," *The Scientist*, 13[10]:21 (May 10, 1999), a
10 copy of which is attached as Exhibit D ("Wilkinson"), the interaction between the antibodies used
11 by Seethala and the phosphotyrosine involves electrostatic interactions. Wilkinson details the
12 antibodies used in his study. These include antibodies designated as PY20, PY54, and PY69
13 (Wilkinson at MDC 0009057). Seethala uses these same antibodies in his study (Seethala at MDC
14 0009043).

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14 **The IMAP Assay Only Measures the Fluorescence Polarization of a Mixture Once**

15 29. The product inserts that accompany the IMAP assay kits, on which Caliper relies, do not
16 instruct users to perform either a “comparing” step or a “monitoring” step. Langer Ex. C. Instead,
17 they instruct users to make only one measurement of fluorescence polarization for a given sample,
18 after the reaction has been incubated to permit it to finish. *Id.* at MDC 0024903 (steps 9-10).

19 30. In the IMAP assay, only the fluorescent polarization of the second mixture is measured –
20 no measurement is made of a first mixture before ATP is added to it, so there is no “before and
21 after” comparison made. Langer Ex. C at MDC 0024902-03 (steps 1-11). A comparison step is,
22 therefore, completely lacking from the instructions that accompany the IMAP assay kits.

23 31. Moreover, the IMAP assay instructions do not tell users to monitor the reaction – they
24 do not tell users to make multiple measurements of the fluorescence polarization of a reaction
25 mixture. The IMAP assays do not measure a *difference* in fluorescence polarization – nor do they

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27 fluo
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1 make other multiple measurements of fluorescence polarization. Instead, they only measure the
2 fluorescence polarization once, at the very end, after the reaction has been allowed to complete.
3 Langer Ex. C at MDC 0024903 (steps 9-10).

4 **Conclusions**

5 32. If the Caliper patent claims – claim 1 of the '774 patent and claim 1 of the '141 patent --
6 require charge-dependent, non-specific interactions, then it is my opinion that they are not infringed
7 by the use of the IMAP assay, because the IMAP assay does not use charge-dependent, non-specific
8 interactions. Instead, the IMAP assay uses coordinate covalent binding. This is illustrated with the
9 figures in Exhibit I.

10 33. Moreover, if the Caliper patent claims require making more than one measurement of
11 the fluorescence polarization of a mixture, then it is my opinion that users who follow the
12 instructions that accompany the IMAP assay kits do not infringe the claims, because those
13 instructions only tell users to make one single measurement of the fluorescence polarization of a
14 given mixture.

15 34. In addition, every element of claims 1 of the '774 and '141 patents is present in the
16 Seethala method. I understand that Seethala is prior art, and that if every element of claims is
17 present in a prior art reference, the claims are invalid, and thus I conclude claims 1 of the '774 and
18 '141 patents are invalid. This is illustrated with the figures in Exhibit I.

19 35. I declare under penalty of perjury under the laws of the United States of America that the
20 foregoing is true and correct to the best of my knowledge.

21 Executed this 30 day of March, 2003 at Houston, Texas.

22 
23 _____
24 Andrew R. Barron
25 _____
26 _____
27 _____
28 _____

Roger S. Borovoy (# 031209)
David J. Miclean (# 115098)
Kurtis D. MacFerrin (# 178006)
Tamara Fraizer (# 215492)
FISH & RICHARDSON P.C.
500 Arguello Street, Suite 500
Redwood City, CA 94063
Telephone: (650) 839-5070
Facsimile: (650) 839-5071

**Attorneys for Defendant and Counter-Claimant
MOLECULAR DEVICES CORPORATION**

UNITED STATES DISTRICT COURT
NORTHERN DISTRICT OF CALIFORNIA
SAN FRANCISCO DIVISION

CALIPER TECHNOLOGIES CORP.,

Case No. C 02-01837 JSW (JL)

Plaintiff,

v.

MOLECULAR DEVICES CORPORATION.

Defendant.

**DECLARATION OF J. RICHARD
SPORTSMAN IN SUPPORT OF
MOLECULAR DEVICES'S OPPOSITION
TO CALIPER TECHNOLOGIES'
MOTION FOR PRELIMINARY
INJUNCTION**

[REDACTED VERSION]

Date: May 16, 2003

Time: 9:00 A.M.

Ctrm: 2

Judge: Honorable Jeffrey S. White

AND RELATED COUNTERCLAIMS.

Roger S. Borovoy (# 031209)
David J. Miclean (# 115098)
Kurtis D. MacFerrin (# 178006)
Tamara Fraizer (# 215492)
FISH & RICHARDSON P.C.
500 Arguello Street, Suite 500
Redwood City, CA 94063
Telephone: (650) 839-5070
Facsimile: (650) 839-5071

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AND RELATED COUNTERCLAIMS.

I, J. Richard Sportsman, declare and state:

1. I am currently an employee of defendant/counterclaimant Molecular Devices Corporation (“MDC”), with the position of Vice President, Assay and Reagent Research and Development. If called as a witness, I could competently testify to the following facts, all of which are within my own personal knowledge.

1 **General Background**

2 2. I am trained as an analytical biochemist. I received a Ph.D. in Analytical Chemistry from
3 the University of Arizona in 1982, and I completed post-doctoral studies in Clinical Immunology at
4 Scripps Clinic and Research Foundation in 1984.

5 3. I have worked at several biotechnology or pharmaceutical companies, including Eli Lilly
6 and Company, SyStemix, Inc., and Terrapin Technologies (now Telik). Each of these companies is
7 or was involved in drug discovery or development of human therapeutics. In my work at these
8 companies, I have acquired special expertise in methods of drug discovery using immunological
9 techniques and fluorescence polarization.

10 4. I have been employed by MDC or L JL Biosystems ("L JL") since August 1998. In
11 August 1998, I joined L JL, which merged with MDC two years later in August 2000. At L JL and
12 MDC, I have been responsible for research on various chemical tests ("assays") and the
13 development of kits that use such tests.

14 5. Further details of my employment history and qualifications are described in my resume,
15 which is attached as Ex. K of this Declaration.

16 **Background for Our Work in Drug Discovery Using Fluorescence Polarization**

17 6. In 1993, I began employment at Terrapin Technologies (now and hereafter Telik) as
18 Director of Molecular Recognition. My research group at Telik was working to develop drug
19 discovery methods suitable for high throughput screening of potential drug compounds. "High
20 throughput screening" or "HTS" is a strategy for drug discovery that came of age in the mid to late
21 1990s. It uses cutting-edge technology, including highly automated laboratory machines, to
22 simultaneously test large numbers of samples – i.e. drug candidates – for potentially desirable
23 characteristics. By screening many samples, we improve our chances of finding new and useful
24 drugs.

25 7. At Telik, we were working to develop drug discovery methods that use a detection
26 technique called "fluorescence polarization." Fluorescence is similar to what makes glow-in-the-
27 dark toys and posters visible after you shine light on them. Things that fluoresce emit light shortly
28 after being "hit" with light. Scientists can attach a molecule that emits fluorescent light – known as

1 a “fluorescent tag” – to a molecule that does not emit fluorescent light, and which would otherwise
2 be invisible. Scientists can then detect the otherwise invisible molecule by the fluorescent “glow” of
3 the tag that was attached to it.

4 8. With the fluorescence polarization technique, scientists measure the polarization of the
5 light that is emitted by the fluorescent tag on a molecule. All light has a polarization direction; it is
6 as fundamental as color. Much like a light beam can be white (mixed colors) or one color, a light
7 beam can have low polarization (many polarization directions) or high polarization (one
8 polarization direction). Natural light, like sunlight, has a low polarization and polarized light, like
9 that which passes through polarized glasses, has a high polarization. When a fluorescent molecule
10 absorbs light with high polarization it naturally emits polarized light. If a fluorescent molecule is
11 stationary for the time between absorbing light and emitting it, one can observe the highly polarized
12 emission. However molecules in solution spin and tumble about. Small molecules tumble quickly.
13 Large ones move more slowly. A group of small, quickly tumbling fluorescent molecules will emit
14 light polarized in many random directions. The net result is light with low polarization. Conversely,
15 a group of large, slowly moving fluorescent molecules tends to emit light with a high polarization.

16 9. The fluorescence polarization of molecules can be used to determine whether there are
17 large or small fluorescent molecules in a sample or mixture: Small fluorescent molecules will have
18 low polarization, while large ones will have high polarization.

19 10. Fluorescence polarization also can be used to determine whether a small fluorescently
20 tagged molecule is bound to a bigger molecule. When the small molecule is bound to a large
21 molecule, it “acts and looks” like a large molecule. That is, it tumbles slowly and has a high
22 polarization. If the small molecule does not bind to the large one, it acts and looks like the small
23 molecule that it is, tumbling fast and having a low polarization. Thus, measuring the fluorescence
24 polarization is one way to determine whether the small molecule is of a kind that could bind to a
25 particular large one. *Low* polarization indicates that the small molecule did *not* bind; *high*
26 polarization indicates that the small molecule *did* bind to the larger one.

27 11. Fluorescence polarization has been known since the 1920s, when scientists first
28 developed the techniques to measure and analyze it. In the 1960s, scientists began using

1 fluorescence polarization to detect the binding of molecules as described above. Beginning in the
2 1980s, fluorescence polarization was used to detect binding in many immunological studies. In the
3 early to mid 1990s, machines were developed that allowed fluorescence polarization to be used for
4 high throughput screening in drug discovery, so that numerous samples could be easily evaluated at
5 the same time.

6 **Background For Our Work on Kinases**

7 12. From 1993 to 1998, while I was at Telik, I and some colleagues were looking for a
8 method that could be used in high throughput screening of chemicals that might affect kinase
9 enzyme activity. Kinases are enzymes that occur naturally in the body, and they are essential for the
10 normal function of cells in our bodies. For example, kinases are important for communication
11 among cells, and their activity can affect cell growth and survival. Abnormal kinase activity may be
12 partly responsible for some cancers, autoimmune diseases, inflammatory diseases (such as asthma),
13 diabetes, neurological disorders and cardiovascular (heart) diseases. Thus, there is considerable
14 interest in finding drugs that inhibit kinase activity (“kinase inhibitors”), and which could therefore
15 be used to control these diseases.

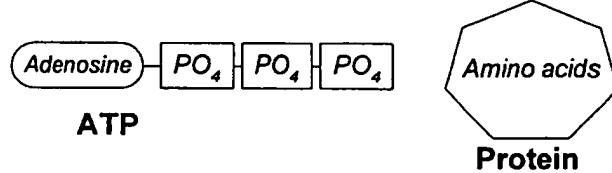
16 13. Although there are many different kinase enzymes in the body, each kinase does one
17 simple thing. It transfers or “moves” a special chemical group called a phosphate (symbolized as
18 PO_4 because it has one phosphorous atom and four oxygen atoms) from one molecule to another.
19 The kinase removes the phosphate from a molecule such as ATP – which stands for adenosine
20 triphosphate. As its names suggests, ATP is a molecule that has *three* phosphates. The kinase
21 removes one of ATP’s phosphates, and attaches that phosphate to another molecule, which could be
22 one of many different proteins. (The removal of a phosphate from ATP transforms it into ADP,
23 which stands for adenosine diphosphate; as its name indicates, ADP has *two* phosphates instead of
24 three, since one phosphate was taken away by the kinase enzyme.) The process of the kinase
25 transferring a phosphate to a protein is called “phosphorylation,” and the molecule receiving the
26 phosphate is then considered “phosphorylated.”

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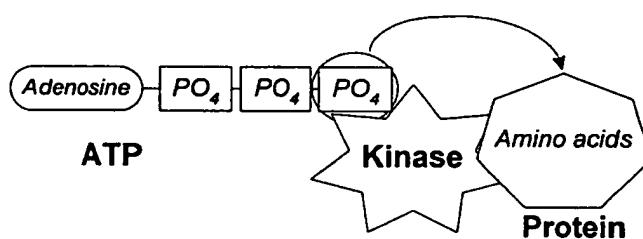
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2 14. Here is a diagram that shows what kinases do:

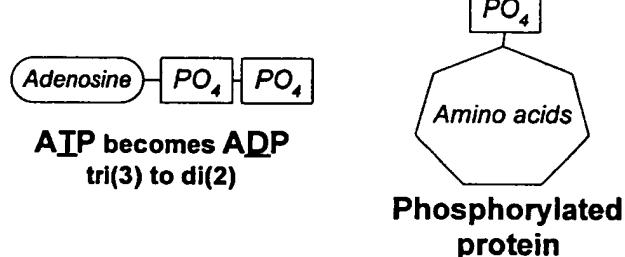
3 Without Kinase Activity



9 The Activity of the Kinase

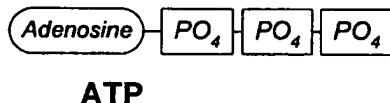


14 The Result of Kinase Activity



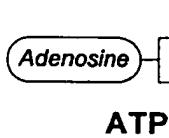
15. In laboratory studies of kinase activity, a “peptide” is typically used instead of a full-length protein. A peptide is just a small piece of a protein. The peptide can be fluorescently tagged, as shown here:

Without Kinase Activity



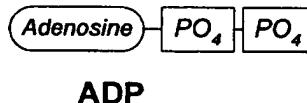
The logo consists of a stylized asterisk (*) above a triangle, with the text "Fluorescently Tagged Peptide" below it.

The Activity of the Kinase



Fluorescently Tagged Peptide

The Result of Kinase Activity



**Fluorescently
Tagged and
Phosphorylated
Peptide**

16. There are two kinds of kinases: “tyrosine kinases” and “serine/threonine kinases.” They differ in terms of where on the protein or peptide the phosphate is attached. Tyrosine kinases attach a phosphate group to a “tyrosine” (one of the 20 amino acids that make up all proteins and peptides), turning it into a “phosphotyrosine.” Serine/Threonine kinases attach a phosphate group to either a “serine” or a “threonine” (two other of the 20 amino acids). Thus, there are two kinds of phosphorylated proteins or peptide: those that have a phosphotyrosine, and those that have a phosphoserine or a phosphothreonine.

Development of our Fluorescence Polarization Antibody Method to Test Tyrosine Kinases

17. While at Telik, from 1993 to 1998, I and my colleagues developed what has been called a "direct" method to screen for chemicals or compounds that inhibit tyrosine kinase activity. The method used fluorescence polarization and antibodies.

18. Antibodies are a special kind of molecule produced by the immune systems of humans and animals. An antibody binds to some specific part of some other molecule. For example, an antibody can bind only to a specific region of a protein. We used an antibody that would bind to a single amino acid in a protein – namely, tyrosine. However, our antibody would bind to tyrosine *only if* that tyrosine had been phosphorylated by a tyrosine kinase (i.e. only if the tyrosine has a phosphate attached to it).

19. We used this “phosphotyrosine antibody” and fluorescence polarization to determine whether or not a phosphate had been transferred by a kinase to a peptide. If a phosphate had been transferred by the kinase, the antibody would bind to the phosphorylated peptide, which would then “act and look” like a larger molecule. That is, it would tumble slowly and have a high polarization. If the kinase enzyme was inhibited, and the peptide was not phosphorylated, the peptide would act and look like the small molecule that it is, tumbling fast and having a low polarization.

20. Because antibodies are expensive, we also devised a variation on this fluorescence polarization assay method, called a “competition” method, that requires fewer antibodies and is therefore less expensive than the “direct” method described above. We filed a patent application on our fluorescence polarization / antibody methods for testing kinase activity. The patent rights were assigned to Telik.

21. The direct method that we devised at Telik is similar to the fluorescence polarization kinase assay methods described by R. Seethala and R. Menzel in their 1997 paper entitled A Homogenous, Fluorescence Polarization Assay for Src-Family Tyrosine Kinases. A true and correct copy is attached hereto as Ex. L. All of these methods use an antibody to bind to a phosphorylated peptide or protein so that it can be detected by a measurement of fluorescence polarization.

Analyst is a large, automated instrument for doing high-throughput screening. It can measure

1 different types of physical features of a sample. One of the things that it can measure is fluorescence
2 polarization. The Analyst can also measure fluorescence intensity, time-resolved fluorescence,
3 luminescence, and absorbance. We used the Analyst to monitor fluorescence polarization in the
4 competition methods noted above. It worked well.

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8 LJL released the
9 first such kit, the "TKX" kit, in April 1998. This and similar kits have been sold by MDC since the
10 LJL - MDC merger in August 2000.

11 **The Next Step: Our Invention of IMAP™**

12 24. Fluorescence polarization methods that use proteins such as antibodies to monitor kinase
13 activity have a number of undesirable features. First, antibody binding is a complicated interaction,
14 involving both structural matching of the protein and the antibody, and various types of interactions
15 including electrostatic interactions. Many things can interfere with the binding, including, for
16 example, the acidity and the salt content of a solution. Second, the competition methods have a
17 relatively weak fluorescent signal, so measuring the fluorescence polarization is difficult. If
18 mistakes are too common, the results are simply not meaningful. Third, there are no antibodies that
19 bind generically to phosphoserine or phosphothreonine (regardless of the protein or peptide), so the
20 methods cannot be readily commercialized for use with all serine/threonine kinases – only for
21 tyrosine kinases. Lastly, antibodies are proteins and, like any protein, their structure and function
22 can be completely disrupted by certain conditions, such as heat or acidity, making them difficult to
23 handle and use.

24 25. When I joined LJL in August 1998, I was interested in developing assay methods that
25 could be used to test all kinases (not just the tyrosine kinases), and which would be robust and easy
26 to use.
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1 26. I knew about the specific nature of bonding between phosphate and iron (Fe(III)) for at
2 least two reasons. First, phosphate was commonly used to determine the presence of iron in a
3 solution; it would bind specifically to the iron, and the bound complexes could be detected in
4 various ways. Second, work published in the 1980's by Porath and others described the use of beads
5 coated with iron to bind specifically to phosphorylated peptides and proteins.

6 The work described by Porath is a type of Immobilized Metal Affinity Chromatography
7 – “IMAC” for short. Chromatography is a common laboratory technique. It uses a column packed
8 full of beads. A solution is poured over the beads in the column, and certain components of the
9 solution bind to the beads while others flow through the column. In this way, the components that
10 bind to the beads can be separated from those that do not.

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I then read an article by M. Posewitz and P. Tempst entitled Immobilized Gallium (III)
Affinity Chromatography of Phosphopeptides. A true and correct copy is attached hereto as Ex. M.
This article shows quite clearly that the metal called gallium (Ga(III)) works better than iron in
IMAC of phosphorylated proteins and peptides.

Commercial Development of IMAP™

38.

We later decided to call our methodology

“IMAP™” as a play on “IMAC” – the affinity chromatography technology that had inspired it.

27 IMAC stands for “immobilized metal affinity column,” while IMAP™ is sometimes said to stand
28 for “immobilized metal affinity for phosphate.”

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17 41. I gave a presentation about our IMAP™ technology at the Society for Biomolecular
18 Screening conference in Baltimore, Maryland, September 10-13, 2001. Theo Nikiforov and other
19 Caliper employees were present at the conference and attended my presentation.

20 42. Aside from the PDE kit, MDC's first two IMAP™ kits were released in December of
21 2001. To date, MDC has developed 17 types of kits (other than the PDE kit) that rely on the
22 IMAP™ technology. Two types were released in December 2001 (Akt and SGK), two in March
23 2002 (p38 and MAPKAP-K2), six in June 2002 (fyn, lck, src, PRAK, MSK1, PP2A), and seven in
24 December 2002 (Blk, Lyn, Syk, CHK1, Chk2, ROCKII, and PTP-1B). The kits are identical except
25 for the particular enzyme and substrate that each contains. Each type of kit is named for the
26 particular enzyme that it contains. Most of the kits include kinase. Two kits, PTP-1B and PP2A,
27 contain a type of enzyme called a phosphatase. A phosphatase *removes* a phosphate from a peptide
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1 or protein (in contrast to a kinase, which *adds* a phosphate to a peptide or protein). Because the
2 enzyme and peptide are included, each kit is “ready-to-use.”

3 43. Each IMAP™ kit is available in “explorer” size (enough material for 800 tests) and
4 “bulk” size (enough material for 8,000 tests). Each IMAP™ kit contains the enzyme (i.e. a
5 particular kinase or phosphatase, or PDE), a fluorescently tagged substrate (e.g. a peptide upon
6 which the enzyme can act), and a solution or “buffer” for mixing them. Each kit also contains
7 gallium-coated nanoparticles (the “binding reagent”) and a special solution (the “binding buffer”),
8 which are combined by the user just before use (to make the “binding solution”). ATP must be
9 supplied by the user.

10 44. MDC sells its binding buffer and binding reagent in bulk, without enzymes or peptides,
11 under the Screening Express program and the IMAP™ purchase plan. These programs allow
12 companies who are able to develop their own particular combinations of enzymes and peptides to
13 purchase only the binding reagent that they need in order to use the IMAP™ technology.

14 45. IMAP™ kinase kits (or materials purchased under the “IMAP™ purchase plan” or
15 “screening express” products) are used to screen potential drug candidates as follows. All of these
16 steps happen in each little well of a multi-well (e.g. a 384- or 1536-well) microplate:

- 17 a. First, the kinase and any drug compounds being tested (i.e. as potential inhibitors of
18 the kinase) are mixed together in a well and incubated. The ensuing test will
19 determine whether the drug inhibits the normal function of the kinase, which is to
20 transfer a phosphate from ATP to a peptide, making it a phosphorylated peptide (see
21 diagrams at Paragraphs 14 and 15).
- 22 b. Next, ATP and fluorescently tagged peptide are added to the well and incubated. If
23 the kinase is not inhibited by the drug compound being tested, it will transfer
24 phosphate from the ATP to the fluorescently tagged peptide, making phosphorylated
25 and fluorescently tagged peptide. This phosphorylated peptide will still be small
26 molecules that tumble quickly and emit light with low fluorescence polarization.
27 Thus, the initial un-phosphorylated fluorescently tagged peptide (if there is any left)

and phosphorylated fluorescently tagged peptide are indistinguishable by fluorescent polarization.

- c. To determine if the fluorescently tagged peptide was phosphorylated by the kinase in step b (and therefore whether the kinase was inhibited or not), a gallium-coated nanoparticle is mixed in a binding solution and added to the well. The binding solution stops the kinase reaction.

the larger gallium-coated

nanoparticle will bind to the phosphorylated peptide (that is, it will bind to the phosphate on that peptide). It will not bind to the fluorescently tagged peptide if the kinase was inhibited and not able to “phosphorylate” the peptide. The purpose of binding the phosphorylated peptide to the gallium-coated nanoparticle is to make a small molecule (the fluorescently tagged peptide), having low fluorescence polarization, into a bigger molecule, having high fluorescence polarization.

- d. The last step is to measure the fluorescence polarization of the chemical mixture in the well. If the kinase did its job and transferred the phosphate from the ATP to the peptide (making it into a phosphorylated peptide), the phosphorylated peptide will bind specifically and robustly, with a coordinate covalent bond, to the larger gallium-coated nanoparticle. The binding will create a larger molecule having high fluorescence polarization. The high fluorescence polarization tells the scientist that the drug did *not* work – the kinase was not inhibited. Conversely, if the drug worked and the kinase was inhibited, such that the peptide was not phosphorylated, then the fluorescently tagged peptide will not bind to the gallium-coated nanoparticle. It will remain a mere fluorescently tagged peptide, tumbling quickly and having low fluorescent polarization..

Summary

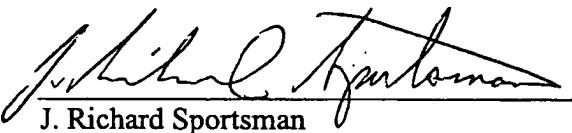
46.

28 I developed IMAP™ with others at MDC based on the available literature, our ideas, and our

1 experiments, and did so well prior to the public issuance of the patents at issue in this case, U.S.
2 Patent Nos. 6,287,774 and 6,472,141 (the '774 and '141 patents). Neither I nor any other scientists
3 at MDC copied or used the system of the '141 and '774 patents in devising our IMAP™ system.

4 47. I declare under penalty of perjury under the laws of the United States of America that the
5 foregoing is true and correct to the best of my knowledge.

6 Executed this 31st day of March, 2003 at Sunnyvale, California.

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8 J. Richard Sportsman
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